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## Palmer LTER: Hydrogen peroxide in the Palmer LTER region: III. Local sources and sinks

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During the austral spring and autumn long-term ecological research (LTER) cruises aboard the R/V Polar Duke (PD92-09, November 1992) and R/V Nathaniel B. Palmer (NBP93-02, March through May 1993), we had an opportunity to investigate selected sources and sinks of hydrogen peroxide ( $H_2O_2$ ) in a variety of antarctic coastal habitats. These measurements constituted one component of our comprehensive study of  $H_2O_2$  dynamics (Karl et al.; Karl and Resing; Resing et al.; Antarctic Journal, in this issue). The potential source terms we evaluated were wet deposition (snow), glacial ice meltwater and land runoff, and *in situ* biological processes; photochemical processes are discussed in a companion paper (Karl and Resing, Antarctic Journal, in this issue). The primary  $H_2O_2$  sink we investigated was bacterial enzymatic activity.

H <sub>2</sub> O <sub>2</sub> wet deposition (snow) in the Palmer LTER study region during austral autumn 1993			
Date	Location	Precipitation H <sub>2</sub> O <sub>2</sub> ª	Sea water H <sub>2</sub> O <sub>2</sub> ª
13 April 1993	64°45'S 64°05'W	432 552	13.6
24 April 1993	67°12.3'S 69°44.5'W	217	10.2
22 April 1993	64°45'S 64°05'W	275	10.1
24 April 1993	67°19.0'S 71°03.6'W	161	8.9
30 April 1993	67°51.3'S 76°00.2'W	55	14.9
7 May 1993	65°55.2'S 65°14.3'W	532 608	15.9
9 May 1993	Humble Island Arthur Harbor	l, 306	6.5

Freshly collected snow samples had consistently elevated concentrations of H<sub>2</sub>O<sub>2</sub> relative to surface sea water (table); the regional average concentration was 349 (±192) nanomoles per liter. These results initially suggest that the atmosphere, through wet deposition, is a local source of H<sub>2</sub>O<sub>2</sub> to surface waters. Based on previous studies, enrichment of H<sub>2</sub>O<sub>2</sub> in marine precipitation was expected (Thompson and Zafiriou 1983), but the values for the LTER study region are lower, by 1-2 orders of magnitude, than rainwaters collected in either the Gulf of Mexico, South Florida, or the Bahama Islands (Zika et al. 1982; Cooper, Saltzman, and Zika 1987). From estimates of the upper water column [0-100 meters (m)] inventories of H<sub>2</sub>O<sub>2</sub> [400-2100 micromole (µmol) per square meter]; Resing et al., Antarctic Journal, in this issue), the mean precipitation rate at Palmer Station [mean of 6.7 millimeters (mm) snow per day during the period November 1992 to January 1993 which is approximately equal to 670 milliliters per square meter per day according to the National Climate Center, Asheville, North Carolina], and our measured dark decay rates of more than 100 µmol per square meter per day (see below), we conclude that wet deposition of  $H_2O_2$  is a weak source term for the LTER study region. Unfortunately, no measurements of H<sub>2</sub>O<sub>2</sub> gas-phase deposition are available.

In addition to the concentrations of  $H_2O_2$  in fresh precipitation, meltwater runoff also contains high levels of  $H_2O_2$  [up to 450 nanomolar (nM)] especially near penguin rookeries. We presently attribute this to an "organic" enrichment and enhancement of  $H_2O_2$  by photoproduction (Karl and Resing, *Antarctic Journal*, in this issue).

Several measurements of the  $H_2O_2$  contents of glacial ice were also made. Floating freshwater ice samples (approximately 10 kilograms each) of unknown origin, were collected during sampling operations in Palmer Basin and Arthur Harbor. Each sample was first rinsed with warm (30°C)  $H_2O_2$ -free distilled water to clean the outer surface, then placed into a clean polyethylene bag and partially melted at room temperature (approximately 20°C). After 10–15 hours, the cold (0°C) meltwaters were collected and analyzed for  $H_2O_2$ . All samples were less than 5 nanomoles (nmol) per kilogram and were consistently lower than the ambient surface sea waters. In contrast to our results, glacial ice samples collected from

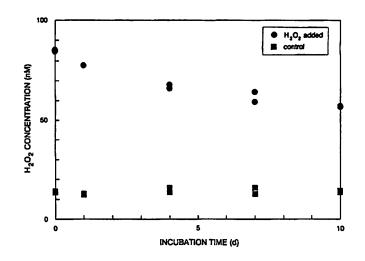
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depths of 100–1,652 meters (m) beneath the antarctic polar plateau at Byrd (79°59'S 120°01'W) and South Pole stations, had  $H_2O_2$  concentrations ranging from 100–500 nmol per kilogram (Neftel, Jacob, and Klockow 1984). The reasons for the differences between these two data sets are not apparent.

Two separate potential biological sources of H<sub>2</sub>O<sub>2</sub> in antarctic coastal waters were also evaluated during the R/V Nathaniel B. Palmer 93-02 cruise. Dark incubations of H<sub>2</sub>O<sub>2</sub>amended surface waters consistently consumed H<sub>2</sub>O<sub>2</sub> (see below), so we concluded that in situ microbial (algae and bacteria) activities comprise a net sink. Nevertheless, two rather serendipitous observations provided evidence for potential biological production of  $H_2O_2$  in Antarctica. During routine collections of krill (Euphausia superba) for physiological experiments by our colleague R. Ross, we observed elevated concentrations of  $H_2O_2$  (up to 250 nM) in sea waters used for short-term (less than 1 hour) containment of dense populations of freshly captured animals and in darkened experimental tanks containing lower population densities (less than 100 adults per cubic meter). We suspect that the source of  $H_2O_2$  in these samples is microbial but as yet have no experimental proof.

The second serendipitous observation was the discovery of elevated H<sub>2</sub>O<sub>2</sub> levels (approximately 1-2 orders of magnitude above ambient surface water depending upon the location) in the R/V Nathaniel B. Palmer's "uncontaminated seawater" system. With the assistance of Chief Engineer D. Munroe, we gained access to the aft centrifugal pump supply (positioned approximately 2 m inboard from the 3-inch diameter intake system in the hull of the ship) which supplies sea water to the main laboratories. The  $H_2O_2$  concentration at the pump was a factor of 2-3 times (up to 20 nM) greater than the surface values collected by Niskin bottles, but was much lower than the H<sub>2</sub>O<sub>2</sub> concentrations in the waters delivered at the laboratory. We conclude that there must be a strong and variable  $H_2O_2$  source within the plumbing of this stainless steel (type 316)/carbon steel (ASTM-A53) sea-water delivery system, despite the fact that the flow rates are large enough for the water temperature at the downstream end to be within 1°C of the incoming sea water. At present, we hypothesize that the source of this  $H_2O_2$  is microbial, rather than chemical.

We conducted several field experiments designed to investigate the nature and potential strength of the microbiological sink for  $H_2O_2$ . Changes in  $H_2O_2$  concentrations were measured over time in sea water incubated in the dark at  $-0.5^{\circ}$ C with and without exogenous  $H_2O_2$  (figure). The  $H_2O_2$ concentrations in unamended sea waters were relatively stable indicating low consumption rates (less than 0.2 nmol  $H_2O_2$  per liter per day). If the sea water is supplemented with  $H_2O_2$  to yield an initial concentration of 85 nM, however, the consumption rate increases to 5 nmol  $H_2O_2$  per liter per day (figure). When the initial  $H_2O_2$  concentration was increased to 1,000 nM, the consumption rate increased to approximately 40 nmol  $H_2O_2$  per liter per day. These results indicate a large potential for dark  $H_2O_2$  catalysis in antarctic surface waters despite a relatively low biomass of living microorgan-



Stability of  $H_2O_2$  in dark incubations at *in situ* temperature. A surface-water sample with and without added  $H_2O_2$  (85 nM) was collected at LTER station 100.140 and incubated in the dark at -0.5°C for 10 days.  $H_2O_2$  concentrations were periodically determined.

isms (0.5–1 µmol living carbon per liter). Nutrient-enriched sea-water cultures (1 gram peptone plus 100 milligrams yeast extract) of heterotrophic bacteria (approximately  $5\times10^6$  cells per milliliter) consumed exogenous  $H_2O_2$  (50 nM) at rates in excess of 50 nmol per liter per hour. The added  $H_2O_2$  was relatively stable (loss rate less than 1 nmol per liter per hour) in sterile-filtered (0.2 µm) treatments.

From our initial investigations, we conclude that both biological and photochemical sources and microbiological sinks of  $H_2O_2$  must be considered in studies of southern ocean  $H_2O_2$  dynamics.

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